# Graphs and structural representations relating to RNA research referenced as 'Dr Fuller'.

### **Contributors**

Fuller, Watson, b.1935

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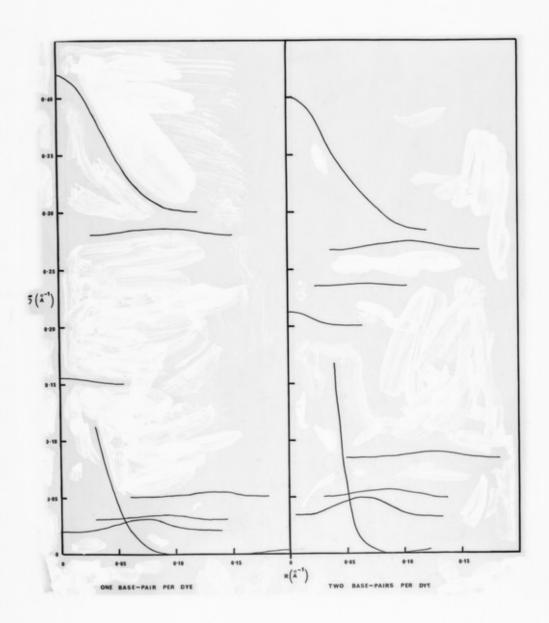
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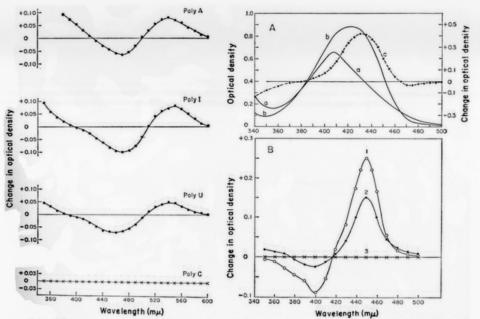
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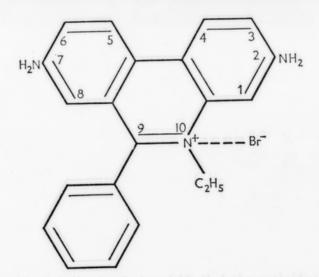




(left). Difference spectra of ethidium solutions read against the same solutions containing polyA, polyI, polyU, and polyC. All measurements were made in 0.1M tris-HCl, pH 7.9. The concentrations of reactants were: ethidium, 40 µg/ml (100 mµmole/ml) (optical density at 480 mµ = 0.54 1-cm light path); polyA, polyI, polyU, and polyC, 400 µg/ml. Fig. 5 (right). A, Effect of Mg\*\* on the spectrum of chromomycin A<sub>2</sub>: Curve a, chromomycin (100 mµmole/ml) in 0.01M tris-HCl, pH 7.4, containing 0.01M NaCl. Curve b, the same, with added MgCl. (0.05M). Curve c, the difference spectrum resulting from the addition of MgCl. B. Effect of Mg\*\* on the interaction of chromomycin with DNA. Difference spectra of chromomycin solutions (100 mµmole/ml) read against the same solutions containing DNA (425 mµmole/ml). Curve I, native calf-thymus DNA. Curve 2, heat-denatured calf-thymus DNA in 0.01M tris-HCl, pH 7.4-0.01M NaCl containing 0.03M MgCl<sub>2</sub>. Curve 3, same as curve I, but without MgCl<sub>2</sub>.

Structures of A, daunomycin (35) ( $R_1$  and  $R_2$  may be as in a or b); B, ethidium bromide (36); C, chromomycin  $A_3$  (37); and D, echinomycin (38). Nogalamycin is similar in structure to daunomycin, possessing a tetracycline-like chromophore linked to an amino sugar. Olivomycin and mithramycin are closely related to chromomycin, differing by minor variations in their chromophores and sugar components (32).

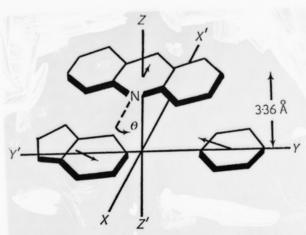
seemendent KNA polymerase (see below) have been used to interpret the in



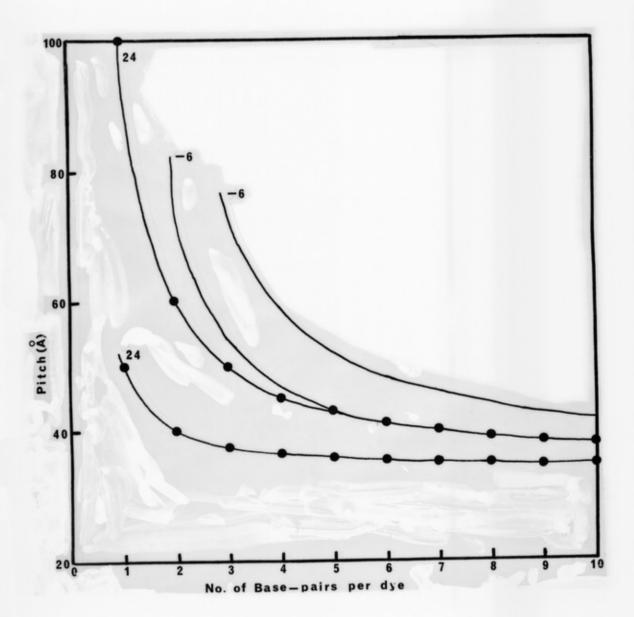
Ethidium bromide: 2:7 diamino-9-phenyl-10-ethyl phenanthridinium bromide.

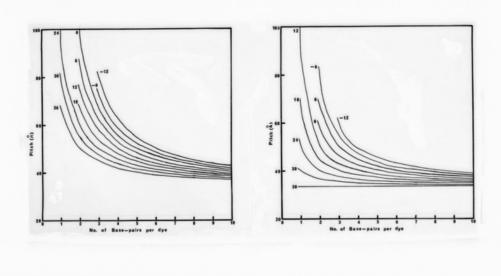
(a) 1. more were propertiones

occus lysodeikticus DNA was prepared from spray-dried cells (California C r Biochemical Research) according to the method of Marmur (1961). After 3

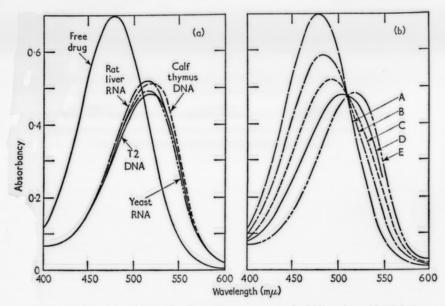


Relative positions of an adenine-thymine base pair and an aminoacridine molecule, as used in the free energy calculations for the intercalated model (I). The skeletons of the molecules are shown and no amino or carboxyl groups are indicated, the ring nitrogen only of the dye being shown. The three dipole moments are represented by short arrows and are not drawn to scale.





 $m\mu$ . This effect is evident as a change in colour from yellow-orange to bright pink. Figure 1(a) shows the results when the nucleic acids are present in excess, that is to say, further additions produce no significant change in the spectrum of the drug. Minor diff



Effect of nucleic acids on the absorption spectrum of ethidium bromide. Solutions contained  $1\cdot25\times10^{-4}$  m-ethidium bromide and the absorbancy was measured using a 1-cm light path. In panel (a) the concentration of each nucleic acid was  $1\cdot5\times10^{-3}$  m. In panel (b) T2 DNA was present at the following concentrations: curve A, zero; B,  $1\cdot5\times10^{-4}$  m; C,  $3\times10^{-4}$  m; D,  $5\times10^{-4}$  m; E,  $1\cdot2\times10^{-3}$  m.

tron spectrum of ethidium bromide. The peak can be seen to shift progressively towards a limit (curve E) which represents the spectrum of the drug in a fully complexed form. All the curves pass through an isosbestic point at 510 m $\mu$ , indicating that they result from the contributions of two forms of ethidium, free and bound, each

ability of the native A-dye system is for a e system over that of a free energy between

a than that of DNA, model adopted must om the binding sites is disrupted (Fig. 5). ecules bound to the ase in  $T'_{\rm m}$ . Stone & endency for acridine efficient is only 1·25. the dye-dye interacharge-dipole and It is these forces, mplex, which could

than for the DNAng that the former DNA-dye complex, tude and direction) charged proflavine

## -dye

The result indicates > AT: AT > TA: GC: GC > CG: GC. one is stabilized the and  $\epsilon'_{ij} = 10$ ,  $\Delta F =$ ace. If the bound dye 4.5 or −4.9 keal, per iated with an A or T ified version of model ongly than AT sites. m measurements of that the relative T > AT:GC > GC:for model I. Hence ding sites. Also, only in  $T_{
m m}$  of the DNAnt samples of DNA

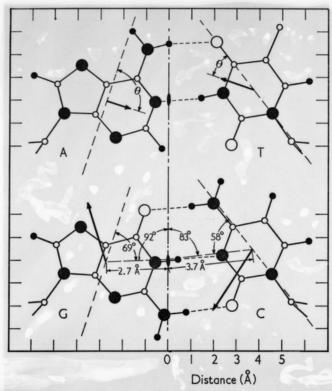
TABLE 3

Comparison of calculated free energy values (in kcal. per repeating unit) for native DNA, model I and model IIa

Repeating unit	De Voe & Tinoco (1962) for native DNA $\epsilon_{ij} = 1 \cdot 0$ $F_{\text{total}}$	Proflavine intercalated, model I $\epsilon_{ij} = 1\text{-}0$		External attachment of proflavine model Ha			
				$\epsilon_{ij}'=10$		$\epsilon'_{ij}=35$	
		$F_{\rm total}$	ΔF	$F_{ m total}$	ΔF	Ftotal	ΔF
CG GC	- 35-7	- 48.3	- 12.6	- 39-6	- 3.9	- 36-9	- 1.2
GC GC	- 19-9	<b>- 48-0</b>	- 28-1	- 23-8	- 3.9	- 21-1	- 1.2
TA CG	- 20.5	- 55-3	- 34.8	- 24-4	- 3.9	- 21.7	- 1.2
AT CG	- 13-7	- 55-3	- 41-6	- 17-6	- 3.9	- 14.9	- 1.2
AT GC	- 12-9	- 55-2	- 42-3	- 16-8	- 3.9	- 14-1	- 1.2
TA GC	- 12.5	- 55-2	- 42-7	- 16-4	- 3.9	- 13.7	- 1.2
GC CG	<b>- 7·5</b>	48-2	- 40-7	- 11-4	- 3.9	- 8.7	- 1.2
TA AT	- 10-6	- 62.7	- 52-1	- 14.5	- 3.9	- 11.8	- 1.2
AT AT	- 10-2	- 62.7	- 52.5	- 14-1	- 3.9	- 11-4	- 1.2
AT TA	- 3-4	- 62-6	- 59-2	<b>- 7·3</b>	- 3.9	- 4.6	- 1.2

The repeating unit shown is for DNA. Those for the DNA-proflavine complexes are those indicated in Fig. 3 with the appropriate base pairs. See text for explanation of the values chosen for  $\epsilon_{ij}$  and for  $\epsilon'_{ij}$ .

of the intercalated model I. For this model, strong binding would be predicted, on the basis of the results in Table 3, up to r=0.5, whereas experimentally it is found that



Atom positions in the AT and GC base-pairs of DNA. The figure is in the plane of the bases and of the dyad axis (— - —) and is perpendicular to the helix axis. The calculated dipole moment of each base is drawn to scale, with the positive end located at the position of the point dipole used in the energy calculations. The dipole orientation angle  $\theta$  is defined. Atom symbols: 
• nitrogen. • oxygen. • hydrogen. • carbon.

